

The genus *Dorofeyukea* (Bacillariophyta, Stauroneidaceae) in karst environments in the Brazilian Pantanal: with the description of four new species

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Abstract: In a Brazilian karst environment, composed predominantly of carbonate rocks, diatoms were collected in periphytic habitat (epiphyton, epilithon, epipsammon) in lentic and lotic water bodies from the Serra da Bodoquena. This Pantanal region is known for its tourist and scenic value, covering springs and rivers with extensive formation of active limestone tufa. Six species of *Dorofeyukea* were investigated under light and electron microscopy. The genus is characterized by a narrow stauroid fascia surrounded by some irregularly shortened striae, uniseriate slightly radiate striae, areolae circular or rectangular covered internally by a hymenes and apical pseudosepta. Four new species are described and compared with similar taxa *Dorofeyukea bahlsii* sp. nov., *D. bodoquenensis* sp. nov., *D. calcarea* sp. nov., and *D. pantanalensis* sp. nov. The new species are distinguished from each other and from similar taxa of *Dorofeyukea* mainly by the valve contour and shape of the apices. *Dorofeyukea kotschy* (Grunow) Kulikovskiy, Kociolek, Tusset et T. Ludwig and *D. texana* (Patrick) Kulikovskiy et Kociolek were also recorded, described and illustrated in this study.

Key words: calcareous habitat, diatom, new species, South America, stauroid fascia, taxonomy

INTRODUCTION

Dorofeyukea Kulikovskiy, Maltsev, Andreeva, T. Ludwig et Kociolek is a naviculoid genus, characterized by solitary cells, described based on molecular analyses and morphological approach of the valve and the chloroplast. *Dorofeyukea* species share narrow stauroid fascia surrounded by 1–3 irregularly shortened striae, the uniseriate and weakly radiate striae, almost parallel at the apices, the circular or rectangular areolae covered internally by hymenes, and the presence of an apical pseudoseptum at both apices. Partial DNA sequences from the SSU and rbcL loci show that *Dorofeyukea* belongs to the stauroneid diatom clade (KULIKOVSKIY et al. 2019).

Currently, twelve species have been accepted taxonomically (GUIRY & GUIRY 2021) and *Dorofeyukea kotschy* (Grunow) Kulikovskiy, Kociolek, Tusset et T. Ludwig is a cosmopolitan species of the genus, found in Europe (KULIKOVSKIY et al. 2019), Africa (STOYNEVA–GÄRTNER & DESCY 2020), Oceania (JOHN 2020), Asia (LIU et al. 2020), and South America (METZELTIN & LANGE–BERTALOT 1998; RUMRICH et al. 2020). *Dorofeyukea*

kotschy was registered as *Navicula kotschy* (Grunow) in floristic freshwater studies from Brazil (TREMARIN et al. 2009; SILVA et al. 2011). Detailed studies revealing the ultrastructure of *Dorofeyukea* species from Brazil are non-existent.

Navicula grimmei var. *rostellata* Hustedt, *Navicula grimmei* Krasske, and *N. grimmeioides* H.P. Gandhi were recently transferred to *Dorofeyukea* (KULIKOVSKIY et al. 2019; TRÁBERT et al. 2019). The detailed analyses of the type materials of *Dorofeyukea rostellata* (Hustedt) Kulikovskiy et Kociolek and *Dorofeyukea grimmeioides* (H.P. Gandhi) Ács, Buczkó et Ector under light (LM) and scanning electron microscopy (SEM) clarified the identity of the Croatian species. Also, the study corroborate that this group did not belong to *Navicula*, and fits in the main features of the recently proposed genus *Dorofeyukea* (TRÁBERT et al. 2019).

The analysis of environmental variables in the karstic water bodies from Serra da Bodoquena (TUSSET et al. 2017, 2018) revealed similarities (pH 7.2–8.3 and conductivity 320–470 $\mu\text{S}\cdot\text{cm}^{-1}$) with those recorded during the studies of *D. rostellata* in Croatian lakes,

located in calcareous bedrock and to those described for *D. indokotschyi* Kulikovskiy, Maltsev, Andreeva et Kociolek and *D. kotschyi* in Indonesia, as neutral to slightly alkaline water (pH 7.3–7.7) and low conductivity (230–240 $\mu\text{S}\cdot\text{cm}^{-1}$) (TRÁBERT et al. 2019, KULIKOVSKIY et al. 2019).

We analyzed populations of six species of *Dorofeyukea* from Brazilian freshwater carbonatic karstic environments, located at Serra de Bodoquena Park, describing and illustrating their morphology by light and scanning electron microscopy. We also compare them with similar taxa, and related its distribution with the physical and chemical water variables. Four species were proposed as new taxa.

MATERIAL AND METHODS

Studied area. The Serra da Bodoquena karst (21°05'42" S, 56°42'17" W) is located at the southwest edge of the Pantanal Complex, in Mato Grosso do Sul, a central west region state of Brazil, covering the cities of Bonito, Jardim and Bodoquena, where sampling occurred. The aquatic environments in the region are characterized by crystal clear and carbonated water and the presence of sinkholes is frequent, despite of the predominance of surface drainage (LINO et al. 1984; SALLUN FILHO et al. 2004). Active limestone tufas are formed by the continuous accumulation of calcium carbonate in association with biological activity (moss and algae) along the drainage system (SALLUN FILHO et al. 2009; BOGGIANI et al. 2011).

Sampling. For this study, 16 sampling sites (waterfall, stream, river and lake) were selected in karst water bodies. A total of 28 periphytic samples were collected in the rainy season: one sample in September 2013; 11 in November 2015 (sampling campaign I); and 16 in November 2019 (sampling campaign II), resulting in six epilithic, three episammic and 19 epiphytic samples (Table 1). We registered the following environmental variables during the sampling: water temperature ($^{\circ}\text{C}$), pH, electrical conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$). Also geographical coordinates are given (Table 1). In the 2019 sampling, other variables were analysed according to APHA (2012): Total P ($\text{mg}\cdot\text{l}^{-1}$), Ca ($\text{mg}\cdot\text{l}^{-1}$), Mg ($\text{mg}\cdot\text{l}^{-1}$), and SiO_2 ($\text{mg}\cdot\text{l}^{-1}$) (Table 2).

Diatom analysis. Periphytic diatoms were obtained by scraping different substrates, such as stones, sand grains, specimens of Charophytes, submerged Bryophytes, with a scalpel wrapped in foil. Samples were fixed in 70% alcohol solution and cleaned according to SIMONSEN (1974) and MOREIRA-FILHO & VALENTE-MOREIRA (1981). Slides were prepared with clean diatom valves, included in Naphrax® (R.I. = 1.74), and were analysed using an Olympus BX40 microscope at 1000 \times magnification (N.A. 1.30, planachromat), equipped with Phase Contrast optics and the Olympus DP71 Imaging System. Cleaned samples were placed on aluminum stubs, dried and covered with gold in Balzers Sputtering SCD 030 for ultrastructural analyses of the frustules in the scanning electron microscopy (SEM). The SEM images were performed in a JEOL JSM 6360LV and TESCAN VEGA3 LMU microscopies, operated at 10–15 kV and 5–8 mm work distance. Besides that, part of the cleaned material was air dried on the formvar-coated copper grids (300 mesh) and analyzed under transmission electron microscopy (TEM) using a JEM 1200EXII, operated at 80 kV. Samples and slides

were deposited in the Herbarium of the Universidade Federal do Paraná (UPCB), and Universidade Federal do Rio Grande do Sul (ICN), Brazil. The terminology used in the description of the taxa was based on BARBER & HAWORTH (1981) and ROUND et al. (1990).

RESULTS AND DISCUSSION

Dorofeyukea bahlsii Tuset sp. nov. (Figs 1–14)

Description

LM (Figs 1–8): Valves elliptic–lanceolate with capitate to subcapitate apices. Valve dimensions (n=20): length 24.2–29.2 μm , width 6–7 μm . Prominent pseudosepta. Axial area relatively broad. Raphe filiform. Proximal raphe ends straight, not expanded. Central area slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly two, forming narrow stauroid fascia bowtie-shaped, slightly expanded to the valve margins. Striae radiate, 22–24 in 10 μm . Areolae not discernible in LM.

SEM (Figs 9–14): Externally, striae continuing onto the mantle, radiate at the center becoming parallel towards the ends (Fig. 9). Areolae transapically elongated in the valve face and longitudinally elongated near the apices, 32–36 in 10 μm . Internally, areolae covered by hymenes (see Fig. 13). Internally and externally, the proximal raphe fissures are slightly deflected in the same direction (Figs 11–12). Internally, distinct, large pseudosepta present at each valve apex (Figs 10, 13). External distal raphe endings hook-shaped to the secondary side, deflected in the same direction, continuing onto the mantle (Fig. 14).

Holotype: UPCB collection No. 78228 (here depicted in Fig. 2), deposited in (Paraná Federal University, Botany Department, Brazil).

Paratypes: Brazil. Mato Grosso do Sul: Bonito, Formoso River, waterfall in Sitio Ybirá Pe farm, epiphyton, UPCB 78254; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epilithon, UPCB 78432; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epilithon, ICN 203543; Mato Grosso do Sul: Bonito, Cachoeiras Park, epilithon, UPCB 78434. Mato Grosso do Sul: Bonito, Cachoeiras Park, epilithon, ICN 203545.

Type locality: Brazil. Mato Grosso do Sul: Bonito, Blue Spring, 20°54'09.7" S, 56°31'48.4" W September 2013, E. Tuset s.n.

Habitat: periphytic (epiphyton, epilithon) in lotic environments.

Etymology: The species name was a tribute to Dr. Loren Bahls for his contribution to the study of diatoms.

Ecology: *Dorofeyukea bahlsii* was very rare in these samples. The populations were found in periphytic habitat (epiphyton, epilithon) of lotic environments. It was found in neutral to alkaline waters (pH 7.6–8), conductivity 387–536 $\mu\text{S}\cdot\text{cm}^{-1}$, water temperature 25–30 $^{\circ}\text{C}$, $\text{N}-\text{NO}_3$ 0.10–0.27 $\text{mg}\cdot\text{l}^{-1}$, Total P 0.01–0.03 $\text{mg}\cdot\text{l}^{-1}$,

Table 1. Sampling data and access number in the Herbarium of the Universidade Federal do Rio Grande do Sul (ICN) and of the Universidade Federal do Paraná (UPCB).

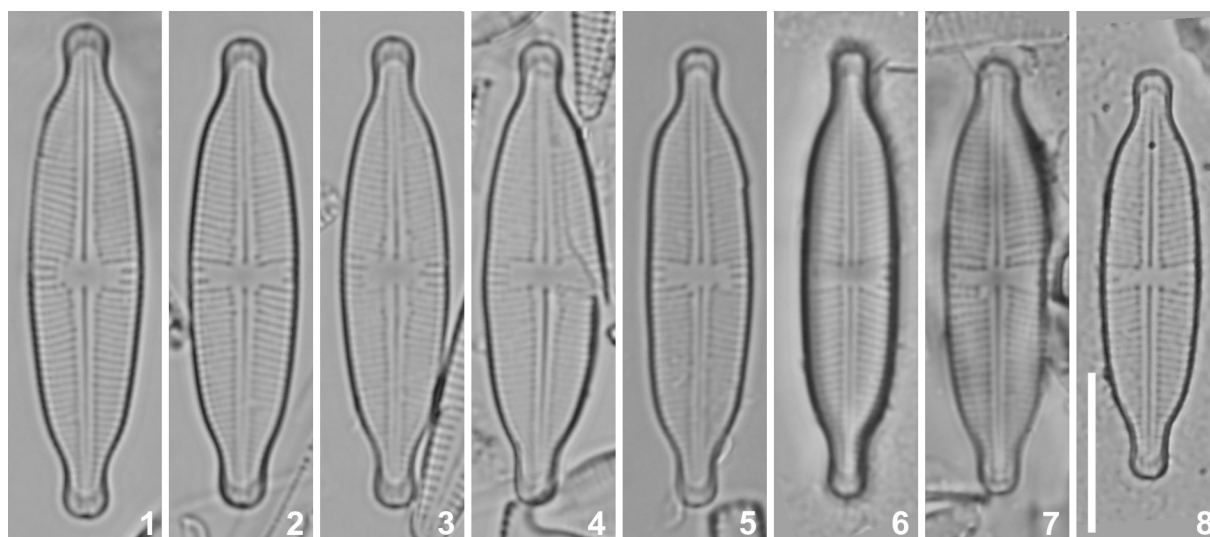
Date	Access UPCB	Access ICN	Locality	City	Geographical coordinates	Sample or substrate	pH	Conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$)	Altitude (m)	Water Tempera- ture (°C)
09/09/2013	78228		Blue Spring	Bonito	20°54'09.7" S, 56°31'48.4" W	Epiphyton / Charophytes	322	...
11/10/2015	76099		Formoso River	Bonito	21°10'17.7" S, 56°27'00.4" W	Epiphyton / Charophytes	7.8	376	285	27
11/10/2015	76100		Sucuri River	Bonito	21°15'12.9" S, 56°33'37.3" W	Epiphyton / Charophytes	7.6	323	325	27
11/10/2015	78233		Olho d'Água River	Jardim	21°26'12.6" S, 56°26'09.3" W	Epiphyton / Charophytes	7.2	320	241	27
11/10/2015	78261		Prata River	Jardim	21°26'08.6" S, 56°25'46.2" W	Epilithon	7.2	320	240	27
11/10/2015	78266		Olho d'Água River	Jardim	21°26'12.6" S, 56°26'09.3" W	Epipsammon	7.2	320	241	27
11/10/2015	78245		Prata River	Jardim	21°26'08.6" S, 56°25'46.2" W	Epiphyton / Charophytes	7.2	320	240	27
11/11/2015	78246		Chapenhina Spring	Bonito	20°50'28.7" S, 56°35'39.8" W	Epiphyton/Bryophytes	7.5	476	308	24
11/11/2015	78248		Chapena waterfall	Bonito	20°49'58.8" S, 56°33'17.3" W	Epiphyton/Bryophytes	7.6	382	308	28
11/12/2015	78254		Formoso River/ Sitio Ybirá Pe waterfall	Bonito	21°09'02.9" S, 56°25'40.7" W	Epiphyton / Charophytes	7.6	387	271	30
11/13/2015	78256		Formoso River/ Ecologic Park	Bonito	21°10'38.1" S, 56°27'08.1" W	Epiphyton / Charophytes	8	376	281	26
11/15/2015	78240		Boca da Onça waterfall	Bodoquena	20°44'23.5" S, 56°44'04.6" W	Epiphyton/Bryophytes	7.3	407	270	27
11/22/2019	78438	203549	Formoso River / Sitio Ybirá Pe waterfall	Bonito	21°09'02.9" S, 56°25'40.7" W	Epiphyton / Charophytes	7.1	411	271	27

Table 1 Cont.

11/22/2019	78453	203564	Bonito River	Bonito	21°08'58.4" S, 56°25'40.3" W	Epiphyton / Charophytes	7.8	422	267	28
11/25/2019	78420	203531	Mimoso River	Bonito	20°59'55.5" S, 56°30'50.1" W	Epiphyton / Charophytes	8	536	246	28
11/25/2019	78431	203542	Prata River	Jardim	21°26'08.6" S, 56°25'46.2" W	Epilithon	7.4	371	240	26
11/25/2019	78432	203543	Mimoso River	Bonito	20°59'55.5" S, 56°30'50.1" W	Epilithon	8	536	246	28
11/25/2019	78440	203551	Misteriosa Lake	Jardim	21°27'30.9" S, 56°27'12.4" W	Epiphyton / Charophytes	8.2	337	254	25
11/25/2019	78443	203554	Olho d'Água River	Jardim	21°26'12.6" S, 56°26'09.3" W	Epiphyton / Charophytes	7.4	338	241	26
11/25/2019	78457	203568	Prata River	Jardim	21°26'08.6" S, 56°25'46.2" W	Epiphyton / Charophytes	7.4	371	240	26
11/25/2019	78456	203567	Olho d'Água River	Jardim	21°26'12.6" S, 56°26'09.3" W	Epipsammon	7.4	338	241	26
11/26/2019	78422	203533	Boca da Onça waterfall	Bodoquena	20°44'23.5" S, 56°44'04.6" W	Epiphyton/Bryophytes	8.6	395	270	26
11/26/2019	78446	203557	Salobra River	Bodoquena	20°44'14.2" S, 56°44'03.9" W	Epilithon	8.1	339	146	29
11/26/2019	78455	203566	Formoso River/ Ecologic Park	Bonito	21°10'38.1" S, 56°27'08.1" W	Epiphyton / Charophytes	8	423	281	27
11/27/2019	78434	203545	Mimoso River/ Cachoeira Park	Bonito	21°00'20.4" S, 56°30'04.7" W	Epilithon	7.7	476	274	25
11/29/2019	78428	203539	Blue Spring	Bonito	20°54'09.7" S, 56°31'48.4" W	Epiphyton / Charophytes	6.7	669	322	25
11/29/2019	78429	203540	Lake Blue Spring	Bonito	20°53'47.8" S, 56°31'36.9" W	Epilithon	7	...	322	26
11/29/2019	78454	203565	Blue Spring	Bonito	20°54'09.7" S, 56°31'48.4" W	Epipsammon	6.7	669	322	25

Table 2. Chemical variables at sampling sites in 2019.

Locality	Ca (mg.l ⁻¹)	Total P (mg.l ⁻¹)	Mg (mg.l ⁻¹)	N-NO ₃ ⁻ (mg.l ⁻¹)	SiO ₂ (mg.l ⁻¹)
Blue Spring	60.60	0.01	39.10	0.10	10.90
Prata River	63.40	0.02	18.90	0.10	21.40
Olho d'Água River	53.60	0.03	19.10	0.10	23.20
Misteriosa Lake	50.40	0.03	18.50	0.10	21.40
Salobra River	47.20	0.01	4.07	0.66	7.87
Boca da Onça waterfall	26.80	0.01	27.60	0.21	11.20
Mimoso River	64.60	0.02	44.60	0.10	17.20
Mimoso River/Cachoeira Park	64.60	0.02	44.60	0.10	17.20
Formoso River /Sitio Ybirá Pe waterfall	29.10	0.03	16.40	0.27	9.72
Formoso River/Ecologic Park	46.90	0.01	12.60	0.27	8.26

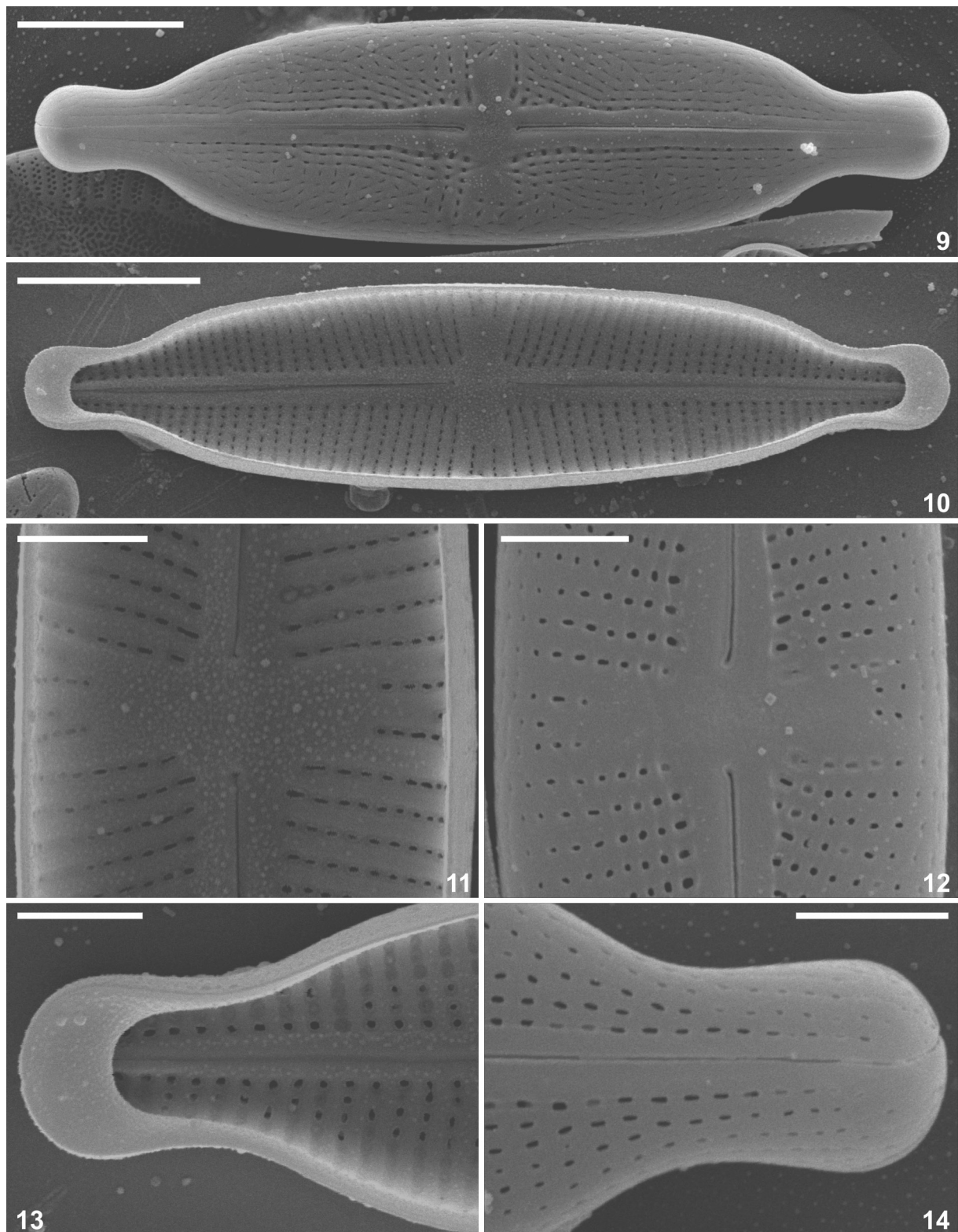
Figs 1–8. *Dorofeyukea bahlsii* sp. nov., LM, valve views: (2) holotype specimen. Scale bar 10 µm.

Ca 29.1–64.6 mg.l⁻¹, Mg 16.4–44.6 mg.l⁻¹, SiO₂ 9.72–17.2 mg.l⁻¹ (Tables 1–2).

Dorofeyukea bahlsii is similar to *Dorofeyukea septata* (Hustedt) J. John in relation to the prominent pseudoseptum, broad axial area, dimensions and radiate striae becoming parallel at the apices. *Dorofeyukea septata* can be distinguished from *D. bahlsii* by the linear–lanceolate to lanceolate valve shape, sub–rostrate to rounded apices, and central area expanded (Table S1). The species was abundant in King Fish Hotspring, in neutral waters (pH 7.2) and high conductivity (1273 µS.cm⁻¹), and in Stevenson Creek, South Australia with alkaline waters (pH 9.1) and low conductivity (207 µS.cm⁻¹) (JOHN 2020).

Dorofeyukea bahlsii have pseudosepta very prominent and quite visible in LM, differing from *D. kotschy*. In addition, *D. bahlsii* has a greater number of areolae in the striae (Table S1) and these are transapically elongated in central area. In *D. bahlsii* the central and axial areas are broader, compared to *D. kotschy* (POTAPOVA 2013; KULIKOVSKIY et al. 2019).

D. indokotschy differs from *D. bahlsii* by the elongated apices, narrow pseudoseptum not discernible in LM, and the less dense striae (20–21 in 10 µm). The population of *D. bahlsii* has elliptic–lanceolate valves, with capitate to subcapitate apices and denser striae, 22–24 at 10 µm (KULIKOVSKIY et al. 2019) (Table S1).



Figs 9–14. *Dorofeyukea bahlstii* sp. nov., SEM: (9–10) external and internal valve view, respectively; (11) internal view of median region of valve showing the narrow fascia slightly expanded to the valve margins with often 1–2 strongly shortened striae and proximal raphe endings; (12) external view of median region showing the proximal raphe endings; (13) internal view of valve apex, note the pseudosepta and areolae occlusion near the apex; (14) external view valve showing the distal raphe ending extending to the mantle. Scale bars 5 μ m (9–12), 2 μ m (13–14).

Dorofeyukea bodoquenensis* Tusset, Tremarin, L.S. Cardoso et T. Ludwig sp. nov. (Figs 15–33)*Description**

LM (Figs 15–26): Valves lanceolate with subrostrate apices. Valve dimensions (n=100): length 11.2–31.8 µm, width 3.6–5.5 µm. Prominent pseudosepta. Axial area relatively broad. Raphe filiform. Proximal raphe endings straight, not expanded. Central area slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly one, forming narrow stauroid fascia, slightly expanded to the valve margins. Striae radiate from the valve center to the apices, 22–26 in 10 µm. Areolae not discernible in LM.

SEM (Figs 27–33): Externally, striae radiate, continuing onto the mantle. Areolae rounded to transapically elongate in the valve face, and longitudinally elongated near the apices, 35–40 in 10 µm. Internally, areolae covered by hymenes (see Fig. 32). Internally proximal raphe ends slightly deflected (Fig. 30). Externally, proximal raphe ends shaped like a tear drop (Fig. 31). Internally, distinct, large pseudoseptum present at each valve apex (Figs 27, 32). In external view the distal raphe end hook-shaped to the secondary side, continuing onto the mantle (Fig. 33).

Holotype: UPCB collection No. 78228 (here depicted in Fig. 19), deposited in (Paraná Federal University, Botany Department, Brazil).

Paratypes: Brazil. Mato Grosso do Sul: Jardim, Chapeninha Spring, epiphyton, UPCB 78246; Mato Grosso do Sul: Bonito, Prata River, epilithon, UPCB 78261; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epiphyton, UPCB 78420; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epiphyton, ICN 203531; Mato Grosso do Sul: Bonito, Blue Spring, epiphyton, UPCB 78428; Mato Grosso do Sul: Bonito, Blue Spring, epiphyton, ICN 203539; Mato Grosso do Sul: Jardim, Prata River, epilithon, UPCB 78431; Mato Grosso do Sul: Jardim, Prata River, epilithon, ICN 203542; Mato Grosso do Sul: Jardim, Misteriosa Lake, epiphyton, UPCB 78440; Mato Grosso do Sul: Jardim, Misteriosa Lake, epiphyton,

ICN 203551; Mato Grosso do Sul: Bodoquena, Salobra River, epilithon, UPCB 78446; Mato Grosso do Sul: Bodoquena, Salobra River, epilithon, ICN 203557; Mato Grosso do Sul: Bonito, Blue Spring, epipsammon, UPCB 78454; Mato Grosso do Sul: Bonito, Blue Spring, epipsammon, ICN 203565; Mato Grosso do Sul: Bonito, Formoso River Ecologic Park, epilithon, UPCB 78455. Mato Grosso do Sul: Bonito, Formoso River Ecologic Park, epilithon, ICN 203566.

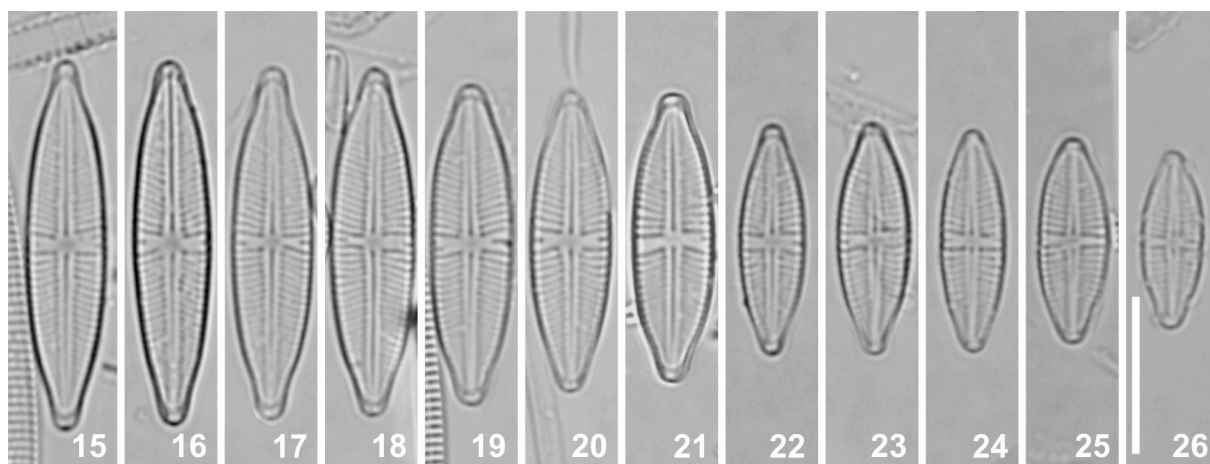
Type locality: Brazil. Mato Grosso do Sul: Bonito, Blue Spring, 20°54'09.7" S, 56°31'48.4" W September 2013, E. Tusset s.n.

Habitat: periphytic (epiphyton, epilithon, epipsammon) in lentic and lotic environments.

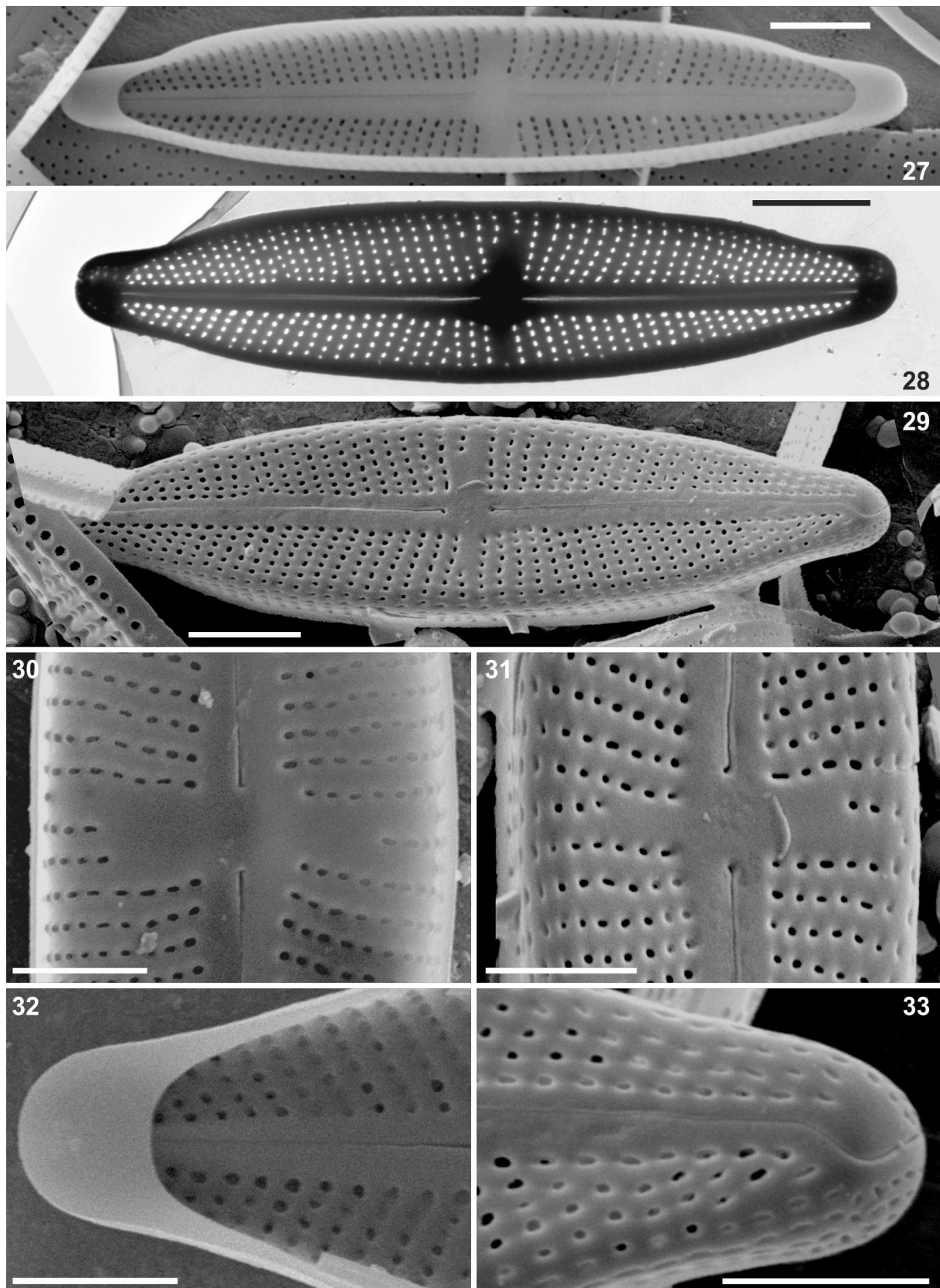
Etymology: The specific epithet refers to the karstic study region of the Serra da Bodoquena National Park.

Ecology: *Dorofeyukea bodoquenensis* was more abundant in the type material. The population were collected in periphytic habitat (epiphyton, epilithon, epipsammon) in lentic and lotic environments. It was found in neutral to alkaline waters (pH 6.7–8.2), conductivity 320–669 µS.cm⁻¹, water temperature 25–29 °C, N–NO₃⁻ 0.10–0.66 mg.l⁻¹, Total P 0.01–0.03 mg.l⁻¹, Ca 46.9–64.6 mg.l⁻¹, Mg 4.07–44.6 mg.l⁻¹, SiO₂ 7.86–21.4 mg.l⁻¹ (Tables 1–2).

The features used to separate *D. bodoquenensis* from similar species of *Dorofeyukea* are presented in Table S1. *Dorofeyukea septata* is similar to *D. bodoquenensis* in terms of its lanceolate valve contour, striation pattern and prominent pseudoseptum at the valve apex. However, these species differ from *D. bodoquenensis* by the rounded areolae and the striae radiate in the center becoming parallel at the ends. Other differences include the central area that is bowtie-shaped (JOHN 2020). Another similar species, *D. calcarea* is distinguished by the slightly expanded central area towards the valve margins (see Figs 46), and also, by the striae slightly radiate along the valve face and almost parallel at the apices. Moreover, pseudosepta are present in the apices, but not easily visible LM (Figs 34–45).



Figs 15–26. *Dorofeyukea bodoquenensis* sp. nov., LM, valve views: (19) holotype specimen. Scale bar 10 µm.



Figs 27–33. *Dorofeyukea bodoquenensis* sp. nov., (27, 29–33) SEM, (28) TEM: (27) internal valve view; (28) valve overview showing the striation pattern; (29) external valve view; (30) internal view of median region of valve showing the narrow fascia slightly expanded to the valve margins; (31) external view of median region showing the proximal raphe endings; (32) internal view of valve apex, note the pseudosepta and areolae occlusion; (33) tilted valve showing the distal raphe ending extending to the mantle. Scale bars 3 μm (27–29), 2 μm (30–33).

Dorofeyukea calcarea* Tusset, Tremarin, L.S. Cardoso et T. Ludwig sp. nov. (Figs 34–51)*Description**

LM (Figs 34–45): Valves lanceolate, minor shapes elliptical, protracted ends, subrostrate to rostrate. Valve dimensions ($n=70$): length 10.5–22 μm , width 3.9–5.5 μm . Narrow pseudosepta. Axial area narrow, linear. Raphe filiform. Proximal raphe endings straight, not expanded. Central area slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly two, forming narrow stauroid fascia bowtie-shaped, slightly expanded to the valve margins. Striae radiate, 20–24 in 10 μm . Areolae not discernible in LM.

SEM (Figs 46–51): Externally, striae continuing onto the mantle, radiate at the center becoming parallel towards the ends. Areolae rounded to transapically elongated in the valve face and longitudinally elongated near the apices, 30–35 in 10 μm . Internally, areolae covered by hymenes (Fig. 51). Externally and internally, the proximal raphe fissures are slightly deflected in the same direction (Figs 48–49). In external view the distal raphe end hook-shaped to the secondary side, continuing onto the mantle (Fig. 50). Internally, distinct narrow pseudoseptum present at each valve apex (Figs 47, 51).

Holotype: UPCB collection No. 78233 (here depicted in Fig. 36), deposited in (Paraná Federal University, Botany Department, Brazil).

Paratypes: Brazil. Mato Grosso do Sul: Bodoquena, Boca da Onça waterfall, epiphyton, UPCB 78240; Mato Grosso do Sul: Jardim, Prata River, epiphyton, UPCB 78245; Mato Grosso do Sul: Bonito, Chapena waterfall, epiphyton, UPCB 78248; Mato Grosso do Sul: Bonito, Formoso River, waterfall in Sítio Ybirá Pe, epiphyton, UPCB 78254; Mato Grosso do Sul: Bonito, Formoso River in Ecologic Park, epiphyton, UPCB 78256; Mato Grosso do Sul: Jardim, Olho d'Água River, epipsammon, UPCB 78266; Mato Grosso do Sul: Bodoquena, Boca da Onça waterfall, epiphyton, UPCB 78422; Mato Grosso

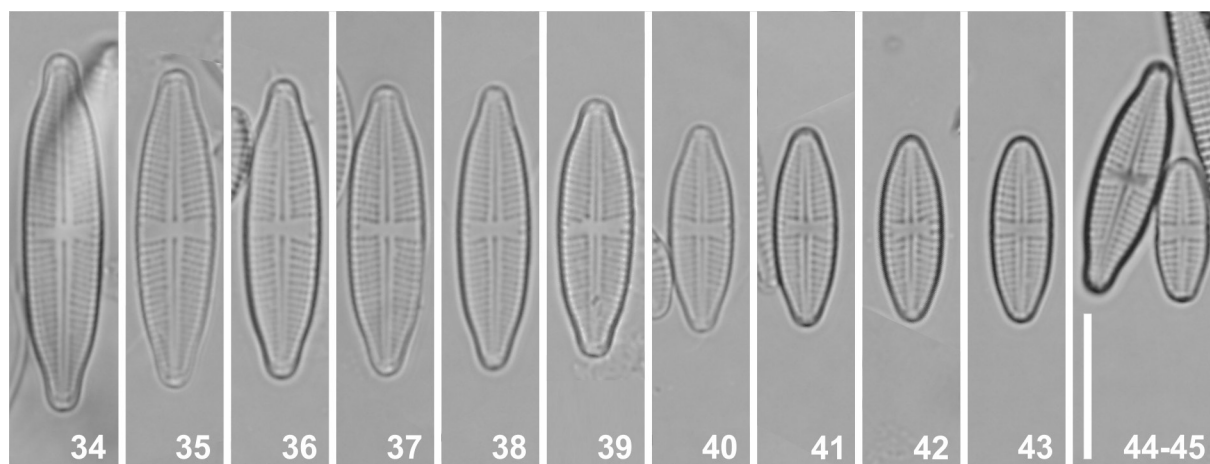
do Sul: Bonito, Blue Spring, epiphyton, UPCB 78428; Mato Grosso do Sul: Bonito, Blue Spring, epilithon, UPCB 78429; Mato Grosso do Sul: Bonito, Blue Spring, epilithon, ICN 203540; Mato Grosso do Sul: Jardim, Prata River, epilithon, UPCB 78431; Mato Grosso do Sul: Jardim, Prata River, epilithon, ICN 203542; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epilithon, UPCB 78432; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epilithon, ICN 203543; Mato Grosso do Sul: Bonito, Cachoeiras Park, epilithon, UPCB 78434; Mato Grosso do Sul: Bonito, Cachoeiras Park, epilithon, ICN 203545; Mato Grosso do Sul: Bonito, Formoso River, waterfall in Sítio Ybirá Pe, epiphyton, UPCB 78438; Mato Grosso do Sul: Bonito, Formoso River, waterfall in Sítio Ybirá Pe, epiphyton, ICN 203549; Mato Grosso do Sul: Jardim, Olho d'Água River, epiphyton, UPCB 78443; Mato Grosso do Sul: Jardim, Olho d'Água River, epiphyton, ICN 203554; Mato Grosso do Sul: Bonito, Bonito River, epiphyton, UPCB 78453; Mato Grosso do Sul: Bonito, Bonito River, epiphyton, ICN 203564; Mato Grosso do Sul: Jardim, Prata River, epiphyton, UPCB 78457; Mato Grosso do Sul: Jardim, Prata River, epiphyton, ICN 203568.

Type locality: Brazil. Mato Grosso do Sul: Jardim, Olho d'Água River, 21°26'12.6" S, 56°26'09.3" W, November 2015, E. Tusset s.n.

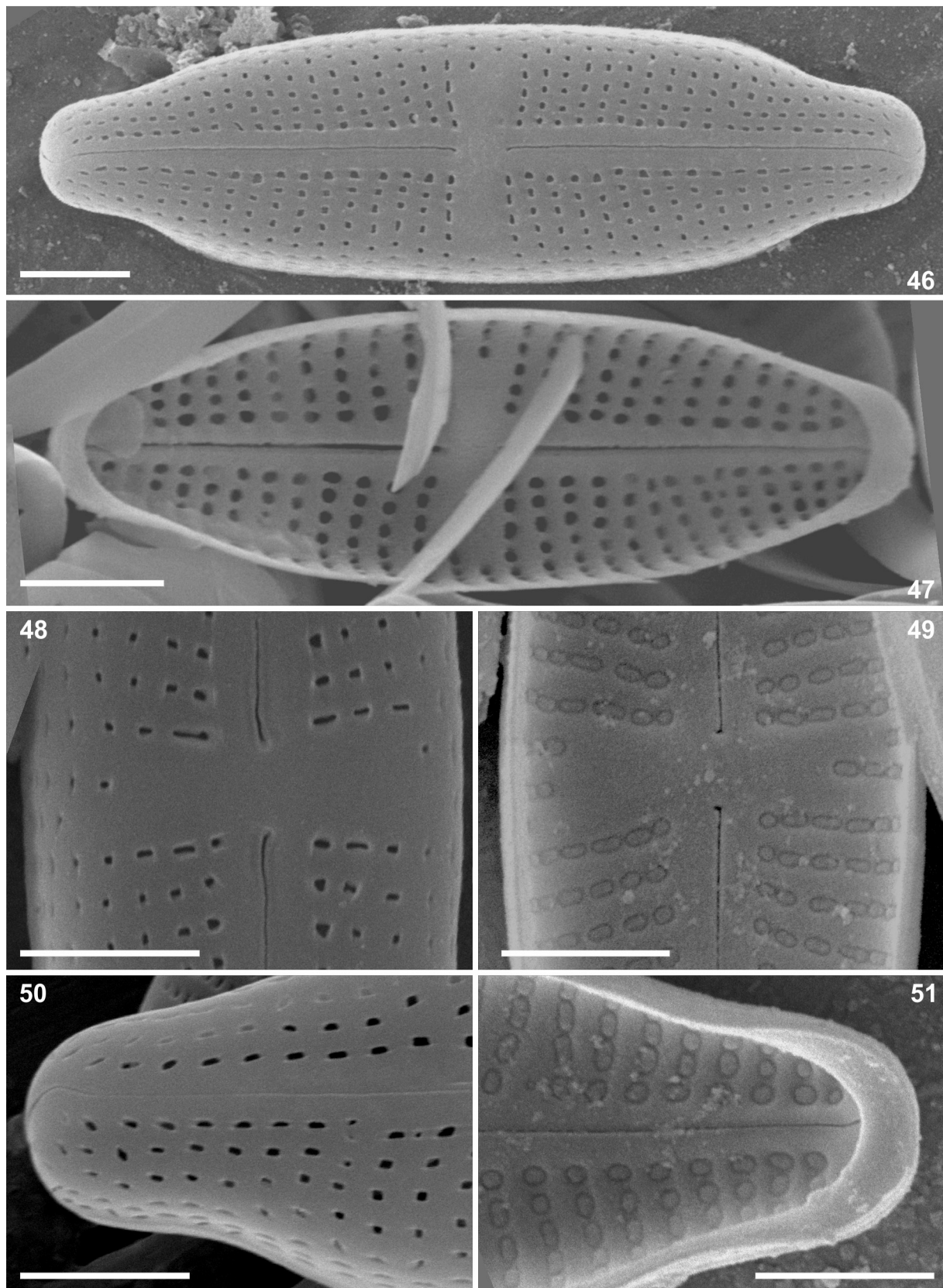
Habitat: periphytic (epiphyton, epilithon, epipsammon) in lentic and lotic environments.

Etymology: The specific epithet refers to the occurrence of the taxon in carbonated waters from the karst landform of the Bodoquena plateau.

Ecology: *Dorofeyukea calcarea* was more abundant in the type material. The population were collected in periphytic habitat (epiphyton, epilithon, epipsammon) in lentic and lotic environments. It was found in neutral to alkaline waters (pH 6.7–8.6), conductivity 320–669 $\mu\text{S}\cdot\text{cm}^{-1}$, water temperature 25–29 °C, $\text{N}-\text{NO}_3^-$ 0.10–0.27 $\text{mg}\cdot\text{l}^{-1}$, Total P 0.01–0.03 $\text{mg}\cdot\text{l}^{-1}$, Ca 26.8–64.6 $\text{mg}\cdot\text{l}^{-1}$, Mg 12.6–44.6 $\text{mg}\cdot\text{l}^{-1}$, SiO_2 8.26–23.2 $\text{mg}\cdot\text{l}^{-1}$ (Tables 1–2).



Figs 34–45. *Dorofeyukea calcarea* sp. nov., LM, valve views: (36) holotype specimen. Scale bar 10 μm .



Figs 46–51. *Dorofeyukea calcarea* sp. nov., SEM: (46–47) external and internal valve view, respectively; (48) external view of central area showing the striation pattern and the proximal raphe endings; (49) internal view of central area showing the narrow fascia; (50) external view of valve apex showing the distal raphe ending; (51) internal view of valve apex, note the pseudosepta. Scale bars 2 μm.

Dorofeyukea calcarea is similar to *D. grimmeioides* and *D. rostellata* in relation to lanceolate valves outline, central area and raphe ends. However, these species differ from *D. calcarea* by having larger valves, radiate striation pattern throughout the valve, lower densities of striae and areolae (Table S1). In addition, *D. grimmeioides* has capitate or rostrate apices, smaller and more elliptical central area with elongate striae. *Dorofeyukea calcarea* has uniform areolae, which is difficult to be observed in LM (Figs 34–45). The central area of *D. rostellata* is transapically oval outside and fairly irregular inside, and areolae can easily be counted in LM. This species has rostrate apices and internal distal raphe ends straight, terminating in a small helictoglossa near the apices (TRÁBERT et al. 2019). *D. Dorofeyukea calcarea* has a maximum valve width of 5.5 µm, while the minimum valve width of *D. rostellata* is 6.6 µm (HUSTEDT 1937).

***Dorofeyukea kotschy* (Grunow) Kulikovskiy, Kociolek, Tusset et T. Ludwig in KULIKOVSKIY et al. (2019) (Figs 52–68)**

Basionym: *Navicula kotschy* Grunow, GRUNOW (1860): Verh. K. Zool.-Bot. Ges. Wien 10: 538, pl.2: fig.12.

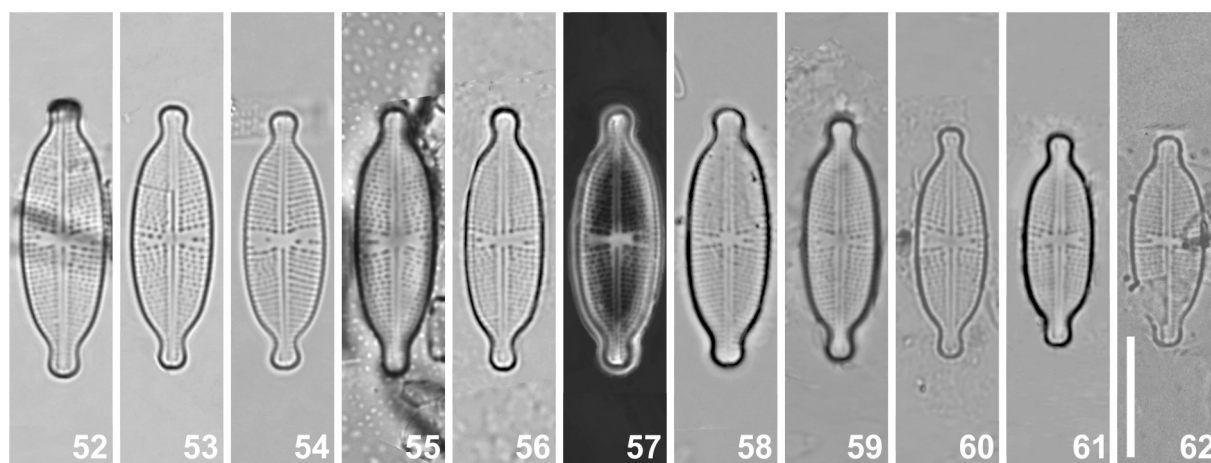
LM (Figs 52–62): Valves elliptic–lanceolate, with rostrate to capitate ends. Valve dimensions (n=35): length 16–23.6 µm, width 5.5–7 µm. Axial area narrow, linear. Raphe filiform. Proximal raphe endings straight, not expanded. Central area slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly two, forming narrow stauroid fascia bowtie-shaped, expanded to the valve margins. Striae radiate, 22–24 in 10 µm. Areolae rounded to transapically elongate, 26–30 in 10 µm.

SEM (Figs 63–68): Externally, striae uniseriate, continuing onto the mantle, radiate at the center becoming parallel towards the ends (Fig. 63). Areolae transapically elongated in the valve face and longitudinally elongated near the apices (Figs 66, 68). Internally, areolae covered by hymenes (see Fig. 67). Proximal raphe fissures internally straight, externally slightly curved (Figs 65–66). Distal raphe endings externally hook-shaped, continuing

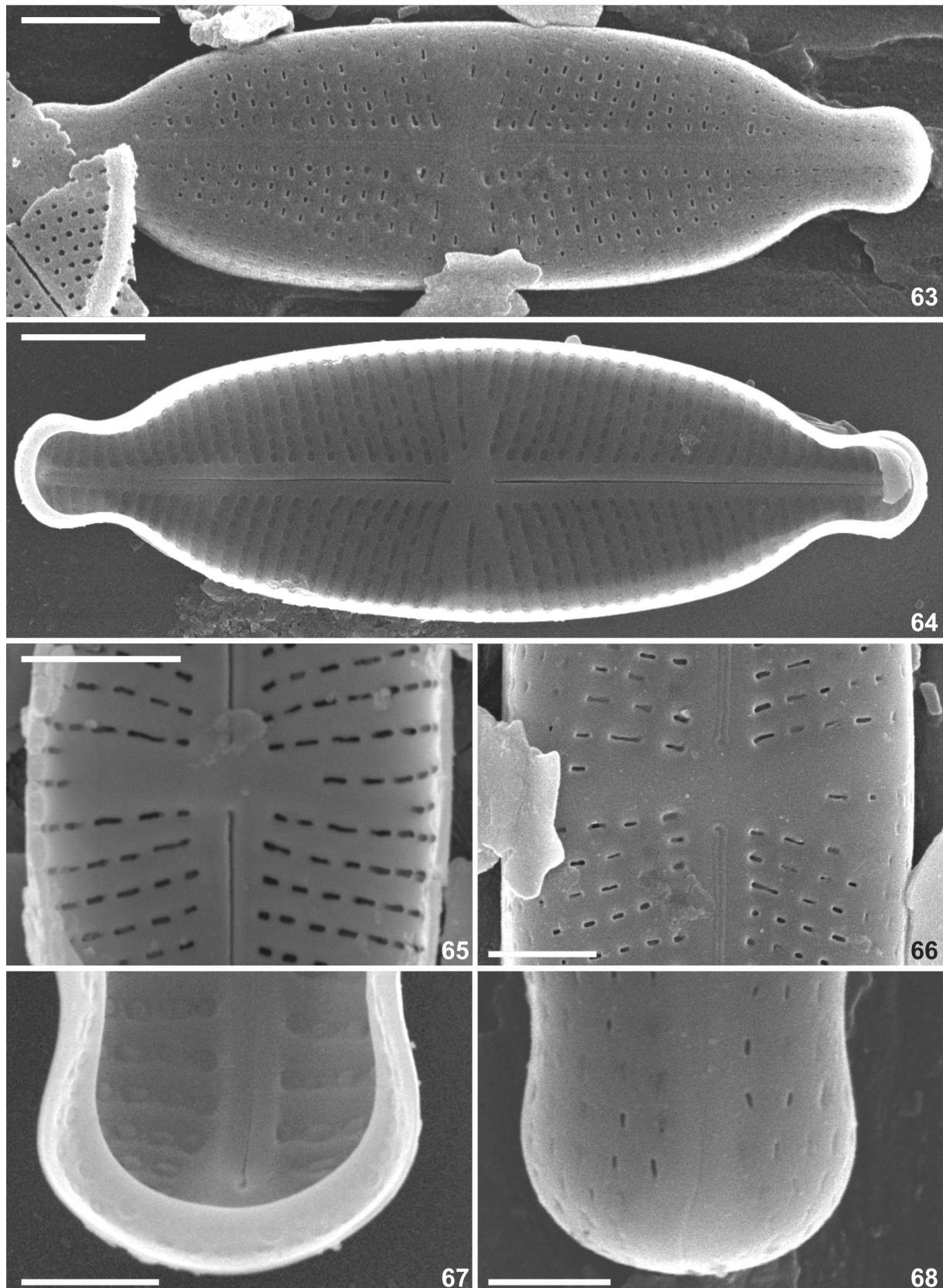
onto the mantle (Fig. 68) and internally ending in a small helictoglossa (Fig. 67). Distinct, but narrow pseudo-septum at each valve apice, ornamented with rounded pores (Figs 64, 67).

Occurrence in samples: BRAZIL. Mato Grosso do Sul: Bonito, Formoso River Ecologic Park, epiphyton, UPCB 76099; Mato Grosso do Sul: Bonito, Sucuri River Ecologic Park, epiphyton, UPCB 76100; Mato Grosso do Sul: Jardim, Olho d'Água River, epiphyton, UPCB 78233; Mato Grosso do Sul: Bodoquena, Boca da Onça waterfall, epiphyton, UPCB 78240; Mato Grosso do Sul: Jardim, Chapeninha Spring, epiphyton, UPCB 78246; Mato Grosso do Sul: Bonito, Chapena waterfall, epiphyton, UPCB 78248; Mato Grosso do Sul: Bonito, Formoso River in Ecologic Park, epiphyton, UPCB 78256; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epiphyton, UPCB 78420; Mato Grosso do Sul: Bonito, waterfall in Mimoso River, epiphyton, ICN 203531; Mato Grosso do Sul: Bodoquena, Boca da Onça waterfall, epiphyton, UPCB 78422; Mato Grosso do Sul: Bodoquena, Boca da Onça waterfall, epiphyton, ICN 203533; Mato Grosso do Sul: Jardim, Prata River, epilithon, UPCB 78431; Mato Grosso do Sul: Jardim, Prata River, epilithon, ICN 203542; Mato Grosso do Sul: Bonito, Formoso River Ecologic Park, epilithon, UPCB 78455; Mato Grosso do Sul: Bonito, Formoso River Ecologic Park, epilithon, ICN 203566; Mato Grosso do Sul: Jardim, Olho d'Água River, epipsammon, UPCB 78456. Mato Grosso do Sul: Jardim, Olho d'Água River, epipsammon, ICN 203567.

Ecology: *Dorofeyukea kotschy* was rare in the samples. The population was observed in periphytic habitat (epiphyton, epilithon, epipsammon) in lentic and lotic environments. It was found in neutral to alkaline waters (pH 7.2–8.6), conductivity 320–536 µS.cm⁻¹, water temperature 24–28 °C, N-NO₃⁻ 0.10–0.66 mg.l⁻¹, Total P 0.01–0.03 mg.l⁻¹, Ca 26.8–64.6 mg.l⁻¹, Mg 12.6–44.6 mg.l⁻¹, SiO₂ 8.2–23.2 mg.l⁻¹ (Tables 1–2).



Figs 52–62. *Dorofeyukea kotschy*, LM, (phase contrast in 57), valve views. Scale bar 10 µm.



Figs 63–68. *Dorofeyukea kotschyi*, SEM: (63) external valve view; (64) internal valve view; (65) internal view of median region of valve showing the narrow fascia slightly expanded to the valve margins; (66) external view of median region showing the proximal raphe endings; (67) internal view of valve apex, note the small helictoglossa, pseudosepta and areolae occlusion near the apex; (68) external view of valve apex showing the distal raphe ending extending to the mantle. Scale bars 5 μm (63–64), 2 μm (65–66), 1 μm (67–68).

Dorofeyukea indokotschyi and *D. orangiana* (Patrick) Kulikovskiy et Kocielek have the external areolae openings transapically elongated and longitudinally elongated near the apices, but differ from *D. kotschyi* by valve shape, apices and number of areolae (Table S1).

D. kotschyi was described from the thermal waters of Budapest, Hungary in 1860, and it is now commonly found across the USA (KULIKOVSKIY et al. 2016 a, b). The species has been reported to Lake Upper Killarney and the river near Glencar (pH 6.3) in County Kerry in Ireland (FOGED 1977). *D. kotschyi* was also reported to Dongting Lake, first record of the genus *Dorofeyukea* to China (LIU et al. 2020).

KULIKOVSKIY et al. (2019) recorded the occurrence of *D. kotschyi* in planktonic habitat (240 $\mu\text{S}\cdot\text{cm}^{-1}$, pH 7.6, water temperature 30 °C) somewhat similar to that where we found *D. kotschyi* in Brazil. *Dorofeyukea kotschyi* [as *Luticola kotschyi* (Grunow) Taylor, Harding et Archibald, nom. inval.] has recently been reported occurring in phytoplankton samples for northwest Argentina in high-altitude from mountain lagoon between 400 and 3.000m (TABOADA et al. 2021).

***Dorofeyukea pantanalensis* Tusset sp. nov. (Figs 69–81)**

Description

LM (Figs 69–75): Valves rhombical–lanceolate, acutely rounded apices, not protracted. Valve dimensions (n=20): length 29.1–33.6 μm , width 5.4–7 μm . Prominent

pseudosepta. Broad axial area. Raphe filiform. Proximal raphe endings straight, not expanded. Central area round, slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly two, forming narrow stauroid fascia, slightly expanded to the valve margins. Striae radiate, 22–24 in 10 μm . Areolae not discernible in LM.

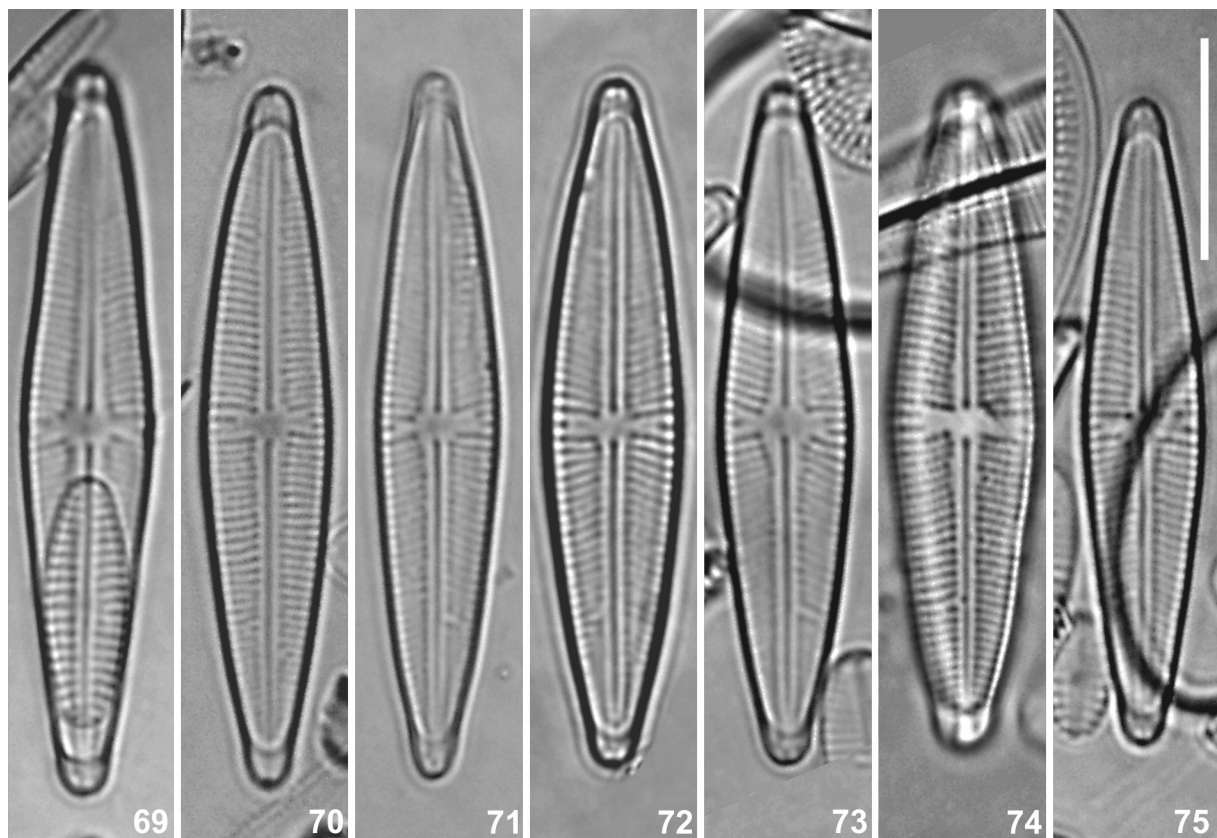
SEM (Figs 76–81): Externally, striae radiate, continuing onto the mantle. Areolae rounded to transapically elongate in the valve face and longitudinally elongated near the apices, 31–35 in 10 μm . Internally, areolae covered by hymenes (see Fig. 80). Internally and externally, the proximal raphe fissures are straight (Figs 78–79). External distal raphe endings hook-shaped to the secondary side, deflected in the same direction, continuing onto the mantle (Fig. 81). Internally, distinct, large pseudosepta present at each valve apex (Figs 77, 80).

Holotype: UPCB collection No. 78233 (here depicted in Fig. 72), deposited in (Paraná Federal University, Botany Department, Brazil).

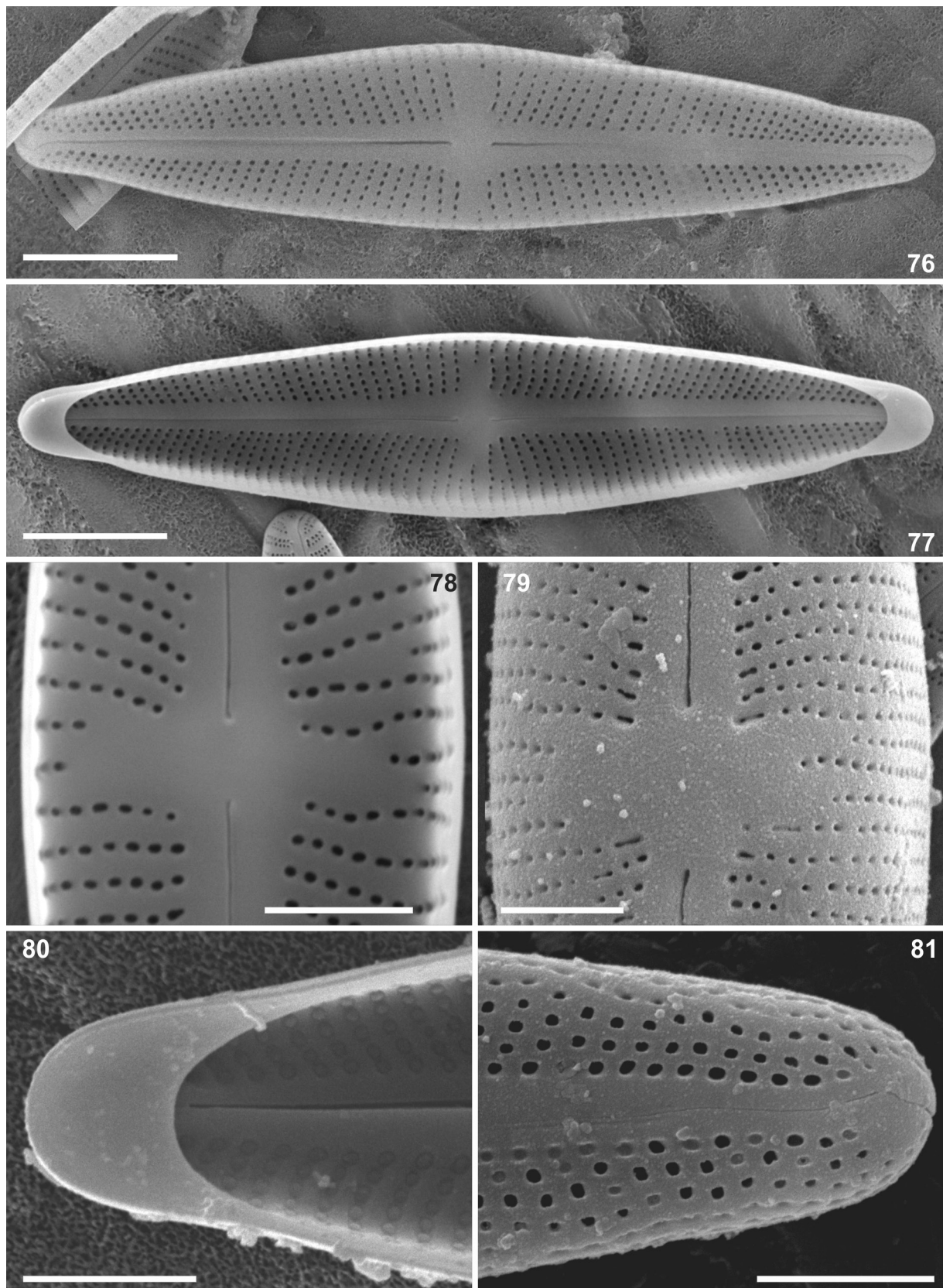
Type locality: Brazil. Mato Grosso do Sul: Jardim, Olho d'Água River, 21°26'12.6" S, 56°26'09.3" W, November 2015, E. Tusset s.n.

Habitat: periphytic (epiphyton) in lotic environments.

Etymology: The species was named in honor to the environment in which the species was found, the Brazilian Pantanal biome.



Figs 69–75. *Dorofeyukea pantanalensis* sp. nov., LM, valve views: (72) holotype specimen. Scale bar 10 μm .



Figs 76–81. *Dorofeyukea pantanalensis* sp. nov., SEM: (76–77) external and internal valve view, respectively; (78) internal view of median region of valve showing the narrow fascia slightly expanded to the valve margins; (79) external view of median region showing the proximal raphe endings; (80) internal view of valve apex, note the pseudosepta and areolae occlusion near the apex; (81) external view of valve apex, showing the distal raphe ending extending to the mantle. Scale bars 5 µm (76–77), 2 µm (78–81).

Ecology: *Dorofeyukea pantanalensis* was very rare in the type material. The population was observed in periphytic habitat (epiphyton) in lotic environment. It was found in neutral waters (pH 7.2), conductivity 320 $\mu\text{S}\cdot\text{cm}^{-1}$, and water temperature of 27 °C (Table 1).

Dorofeyukea pantanalensis is very distinct from any other *Dorofeyukea* species. Rhombic–lanceolate valves, prominent pseudosepta, records of valves with dimensions greater than 30 μm were only reported to *D. ivatoensis* (Metzeltin et Lange–Bertalot) Kulikovskiy et Kociolek, *D. rostellata* and *D. tenuipunctata* (Hustedt) Kulikovskiy et Kociolek, and they can be easily distinguished by the valve shape (Table S1).

Navicula septataeoides Hustedt, rarely reported in the literature, is described and illustrated (pl. 776, fig. 1749) by HUSTEDT (1961–1966) from Lago Alto Branco, Brazil, and it seems to be the taxon most related to *D. pantanalensis*. *Navicula septataeoides* presents prominent pseudosepta and radiate striae along the valve, differing by lacking the stauroid fascia, slightly expanded to the valve margins, and dimensions are much larger, length (73 μm long and 15 μm wide) in relation to *D. pantanalensis*.

D. pantanalensis presents wider valves, rhombic–lanceolate, acutely rounded apices, not protracted, while *D. bodoquenensis* has lanceolate valves with subrostrate ends (Table S1).

***Dorofeyukea texana* (Patrick) Kulikovskiy et Kociolek in KULIKOVSKIY et al. (2019) (Figs 82–99)**

Basionym: *Navicula texana* Patrick, PATRICK (1959): Proc. Acad. Nat. Sci. Philadelphia, 111: 98, pl. 8, fig. 5.

LM (Figs 82–93): Valves lanceolate, with subcapitate to rostrate ends. Valve dimensions (n=30): length 17.5–21.5 μm , width 4–5.6 μm . Narrow pseudosepta. Axial area narrow, linear. Raphe filiform. Proximal raphe endings straight, not expanded. Central area slightly asymmetric due to the varying number of short central striae ranging from one to three, most commonly two, forming narrow stauroid fascia bowtie-shaped, slightly expanded to the valve margins. Striae radiate, 20–24 in 10 μm . Areolae

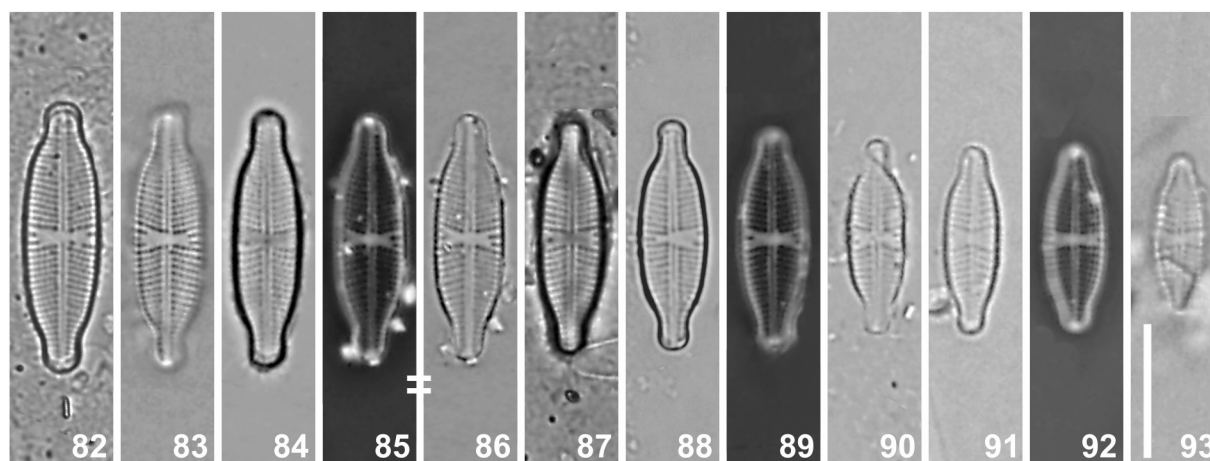
not discernible in LM.

SEM (Figs 94–99): Externally, striae uniseriate, continuing onto the mantle, radiate at the center becoming parallel towards the ends. Areolae transapically elongated in the valve face and longitudinally elongated near the apices, 33–35 in 10 μm . Internally areolae individually covered by hymenes (see Fig. 96). Externally, the proximal raphe fissures slightly deflected in the same direction (Fig. 97). Internally, distinct, narrow pseudosepta present at each valve apex (Fig. 98). External distal raphe endings hook-shaped to the secondary side, deflected, continuing onto the mantle (Fig. 99). Open girdle bands with one row of small pores (Figs 95, 99).

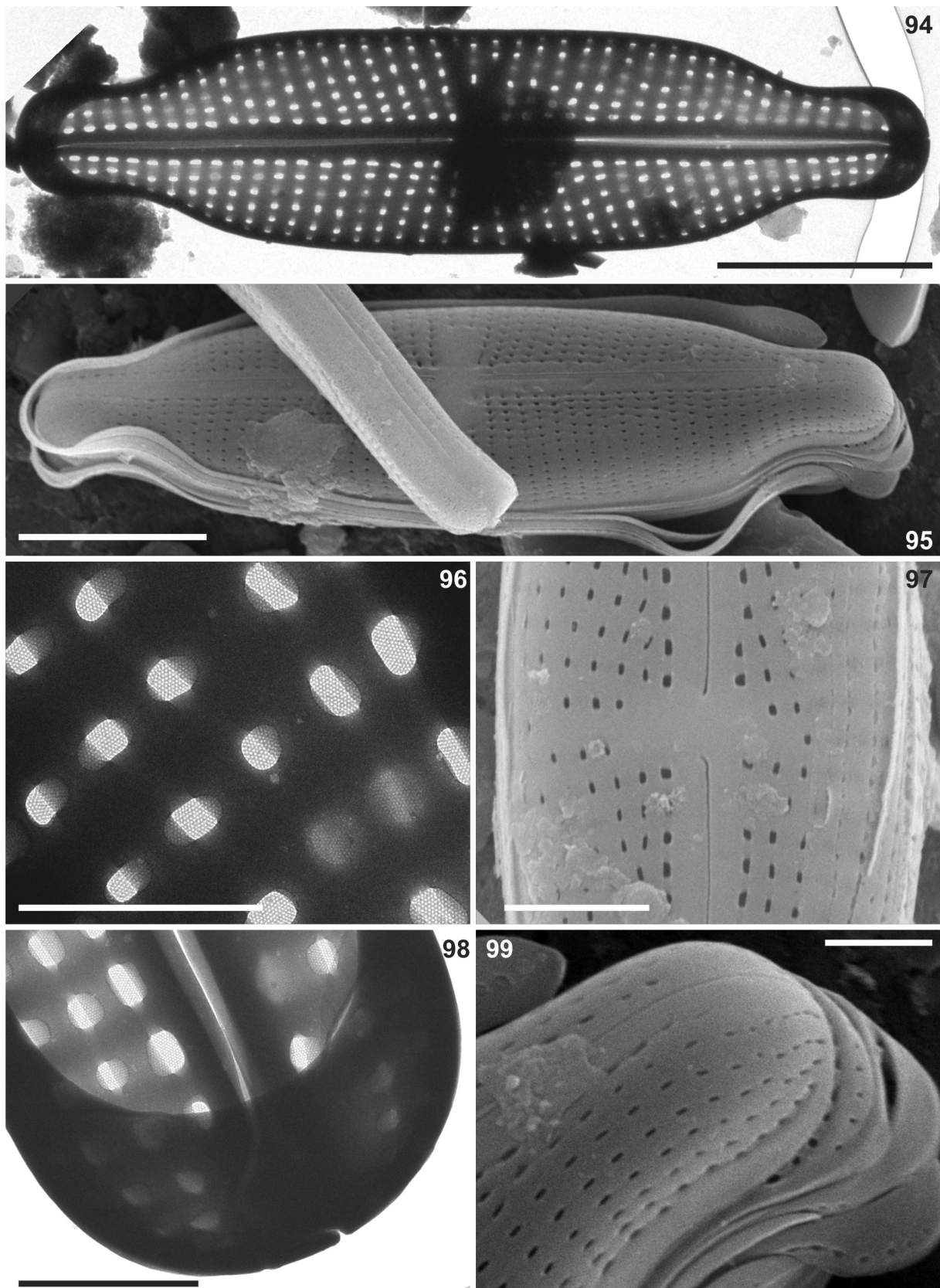
Occurrence in samples: Brazil. Mato Grosso do Sul: Bonito, Chapena waterfall, epiphyton, UPGB 78248; Mato Grosso do Sul: Bonito, Blue Spring, epiphyton, UPGB 78428; Mato Grosso do Sul: Bonito, Blue Spring, epiphyton, ICN 203539.

Ecology: *Dorofeyukea texana* was very rare in the samples. The population was observed in periphytic habitat (epiphyton) in lotic environments. It was found in neutral to alkaline waters (pH 6.7–7.6), conductivity 382–669 $\mu\text{S}\cdot\text{cm}^{-1}$, water temperature 25–28 °C, N- NO_3^- 0.10 $\text{mg}\cdot\text{l}^{-1}$, Total P 0.01 $\text{mg}\cdot\text{l}^{-1}$, Ca 60.6 $\text{mg}\cdot\text{l}^{-1}$, Mg 39.1 $\text{mg}\cdot\text{l}^{-1}$, SiO₂ 10.9 $\text{mg}\cdot\text{l}^{-1}$ (Tables 1–2).

POTAPOVA (2013) investigated the type population of *Navicula texana* Patrick and *N. savannahiana* Patrick, currently included in *Dorofeyukea*. Both species present narrow capitate apices, punctate striae and transverse central area characteristic of *D. kotschyi*, but they differ in width and valve outline. POTAPOVA (2013) also observed a pseudoseptum at each distal valve end of *D. texana*, but she cannot confirm the existence of pseudosepta in *D. savannahiana* due to the rare occurrence of this species in the sample. Based on the illustrations presented by POTAPOVA (2013: Figs 81–87), we concluded that *D. savannahiana* has curve–radiate striae and transapically elongated areolae different from



Figs 82–93. *Dorofeyukea texana*, LM, (phase contrast in 85, 89, 92), valve views. Scale bar 10 μm .



Figs 94–99. *Dorofeyukea texana*, (94, 96, 98) TEM, (95, 97, 99) SEM: (94) valve overview showing the striation pattern; (95) external valve view; (96, 98) detail of areolae occlusion, note the hymens; (97) external view of median region of valve showing the narrow fascia slightly expanded to the valve margins; (98) view of valve apex, pseudosepta; (99) external view of valve and apex showing the girdle bands. Scale bars 5 μ m (94–95), 2 μ m (97), 1 μ m (96, 98–99).

D. texana that have parallel striae and punctiform or apically elongated areolae.

Distribution of *D. texana* (as *Navicula texana*) in North America: Laurentian Great Lakes (STOERMER, KREIS & ANDRESEN 1999), type locality Texas, freshwater with high mineral content, hard water (PATRICK & REIMER 1966), and United States of America (KOCIOLEK 2005, EBERLE 2008). It was also recorded from Asia: Korea (LEE, CHOI & LEE 1995, LEE 2012), Mongolia (DOROFYUK & KULIKOVSKIY 2012), and Russia (Far East) (MEDVEDEVA & NIKULINA 2014).

Dorofeyukea belongs to the stauroneioid diatom forming a single branch clade, together with *Stauroneis* Ehrenberg, *Prestauroneis* Bruder et Medlin, and *Craticula* Grunow, and it is frequently characterized as having a narrow stauroid fascia surrounded by few shortened striae. Also, species are represented by relatively small length cells, not exceeding 40 µm (KULIKOVSKIY et al. 2019; TRÁBERT et al. 2019). *Stauroneis* is a polyphyletic genus and further molecular and morphological investigation are needed (KULIKOVSKIY et al. 2019).

COX & WILLIAMS (2000) stated that in 1894, Cleve had already commented that species with different forms of thickening in the central area were placed in *Stauroneis*, and the stauroids are used as diagnostic characteristic of this genus, and is easily recognized by light microscopy. Based on this supposed feature, the central area can be thickened in various ways and delimited by the shortening of striae (ROUND et al. 1990; KOCIOLEK et al. 2019; COX & WILLIAMS 2000; COX 2001; WADMARE et al. 2022).

Dorofeyukea kotschy was included in *Navicula* before. But, the isodiametric shape of the areolae is not characteristic of the apically elongated areolae of *Navicula* sensu stricto, as well as the pseudoseptum; therefore, these species should be placed in another genus (POTAPOVA 2013). The study of scanning electron microscopy (SEM) is often necessary to determine the taxonomic position of species, recent studies on several original materials have transferred some naviculoid species from *Navicula*, *Naviculadicta* Lange–Bertalot in Lange–Bertalot et Moser to another genus, as well as some taxa belonging to *Dorofeyukea*.

Twelve taxa use to be the total diversity of *Dorofeyukea*. We found two of them occurring in Brazilian karst, and four new species unknown so far, increasing the number of species in the genus *Dorofeyukea* to sixteen. The taxa were observed in periphytic habitat (epiphyton, epilithon, epipsammon) in lentic and lotic freshwater environments. The species *D. pantanalensis*, *D. texana*, and *D. bahlsii* were rare in this study, *D. pantanalensis* and *D. texana* occurred in only one sample while *D. bahlsii* in three. *Dorofeyukea bodoquenensis*, *D. calcarea*, and *D. kotschy* occurred in more than ten samples (Table 1).

Investigating diatom diversity occurring in these karst environments around the world, elevated richness is notable in Florida Everglades (LEE et al. 2013),

Croatia (JASPRICA & HAFNER 2005), Slovakia (CZERWIK–MARCINKOWSKA et al. 2018), Bosnia, Herzegovina, Croatia and Montenegro (HAFNER & JASPRICA 2013), Poland (WOJTAŁ 2003), Vietnam (WEIDE 2015), Mexico (NOVELO et al. 2007; IBARRA et al. 2009), Mediterranean (LAI et al. 2019), Spain (DELGADO et al. 2013), Italy (LAI et al. 2016). Some recent studies resulted in the description of new species of diatoms as in Croatia (GLIGORA UDOVIČ et al. 2022), China (YOU et al. 2016; YOU et al. 2019; LOWE et al. 2017), and Brazil (TUSSET et al. 2017, 2018). Therefore, new species proposals in this study were not surprising.

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REFERENCES

- APHA, A.W.W.A. (2012): Standard Methods for the Examination of Water and Wastewater. – 22nd ed. Washington, DC, USA: American Public Health Association.
- BARBER H.G. & HAWORTH E.Y. (1981): A guide to the morphology of the diatom frustule. – Freshwater Biological Association Scientific Publication 44: 1–112
- BOGGIANI, P.C.; TREVELIN, A.C.; SALLUN FILHO, W.; OLIVEIRA, E.C. & ALMEIDA, L.H.S. (2011): Turismo e conservação de tufas ativas da Serra da Bodoquena, Mato Grosso do Sul. Pesquisas em Turismo e Paisagens Cársticas, Campinas, SETur/SBE. – Tourism and Karst Areas 4: 55–63.
- CLEVE, P.T. (1894): Synopsis of the naviculoid diatoms. – Kungliga Svenska vetenskapsakademiens handlingar 26: 1–194.
- COX, E.J. (2001): What constitutes a stauroid? A morphogenetic perspective. – In: JAHN, R.; KOCIOLEK, J.P.; WITKOWSKI, A. & COMPÈRE, P. (eds): Festschrift für H. Lange–Bertalot. – pp. 303–316., A.R.G. Gantner Verlag, Ruggell.
- COX, E.J. & WILLIAMS, D.M. (2000): Systematics of naviculoid diatoms: the interrelationships of some taxa with a stauroid. – European Journal of Phycology 35: 273–82. DOI: <https://doi.org/10.1080/09670260010001735871>
- CZERWIK–MARCINKOWSKA, J.; WRÓBLEWSKI, W.; GRADZIŃSKI, M. & UHER, B. (2018): Diatom species diversity and their ecological patterns on different substrates in two karstic streams in the Slovak Karst. – Journal of cave and karst studies the National Speleological Society 80: 1–16. DOI: <http://dx.doi.org/10.4311/2017MB0124>
- DELGADO, C.; ECTOR, L.; NOVAIS, M.H.; BLANCO, S.; HOFFMANN, L. & PARDO, I. (2013): Epilithic diatoms of springs and springs-fed streams in Majorca Island (Spain) with the description of a new diatom species *Cymboppleura margalefii* sp. nov. – Fottea 13: 87–104. DOI: <http://dx.doi.org/10.5507/fot.2013.009>

- DOROFYUK, N. & KULIKOVSKIY, M. (2012): Diatoms of Mongolia. Proceedings of Joint Russian–Mongolian Complex Biological Expedition, Vol. 59. – 366 pp, Institute of Ecology and Evolution RAN, Moscow.
- EBERLE, M.E. (2008): Recent diatoms reported from the central United States: register of taxa and synonyms. – 88 pp., Department of Biological Sciences, Fort Hays State University, Hays, Kansas.
- FOGED, N. (1977): Freshwater diatoms of Ireland. – *Bibliotheca Phycologica* 34: 1–221.
- GUIRY, M.D. & GUIRY, G.M. (2021): AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.ORG>
- HAFNER, D. & JASPRICA, N. (2013): The composition of epiphytic diatoms (Bacillariophyta) on Charophyceae in Dinaric karstic ecosystems. – *Natura Croatica* 22: 199–204.
- HUSTEDT, F. (1937): Systematische und ökologische Untersuchungen über die Diatomeenflora von Java, Bali, und Sumatra nach dem Material der Deutschen Limnologischen Sunda-Expedition. Teil I. Systematischer Teil, Erste Fortsetzung. – *Archiv für Hydrobiologie*, Suppl. 15: 187–295.
- HUSTEDT, F. (1953): Diatomeen aus der Oase Gafsa in Südtunesien, ein Beitrag zur Kenntnis der Vegetation afrikanischer Oasen. – *Archiv für Hydrobiologie* 48: 145–153.
- HUSTEDT, F. (1961–1966): Die Kieselalgen. – In: RABENHORST, L. (ed.): Kryptogamen-Flora von Deutschland, Österreich und der Schweiz, Band VII (3, Lief. 2) von Dr. Friedrich Hustedt, Bremen, Reprint 1977. – 816 pp., Otto Koeltz Science Publishers, Koenigstein/West Germany.
- IBARRA, C.; TAVERA, R. & NOVELO, E. (2009): Diversidad y estructura de las comunidades de diatomeas del perifiton y el metafiton en un humedal tropical en México. – *Revista Mexicana de Biodiversidad* 80: 763–769. DOI: <http://dx.doi.org/10.22201/ib.20078706e.2009.003.171>
- JASPRICA, N. & HAFNER, D. (2005): Taxonomic composition and seasonality of diatoms in three Dinaric karstic lakes in Croatia. – *Limnologica* 35: 304–319. DOI: <https://doi.org/10.1016/j.limno.2005.08.003>
- JOHN, J. (2020): Diatoms from arid Australia: taxonomy and biogeography. The diatom flora of Australia Schmittgen – Oberreifenberg, Vol. 3. – 578 pp., Koeltz Botanical Books, Königstein.
- KOCIOLEK, J.P. (2005): A checklist and preliminary bibliography of the Recent, freshwater diatoms of inland environments of the continental United States. – *Proceedings of the California Academy of Sciences*. – Fourth Series 56: 395–525.
- KOCIOLEK, J.P.; WILLIAMS, D.M.; STEPANEK, J.; LIU, Q.; LIU, Y.; YOU, Q. M.; KARTHICK, B. & KULIKOVSKIY, M. (2019): Rampant homoplasy and adaptive radiation in pennate diatoms. *Plant Ecology and Evolution*. 152:131–41. DOI: <https://doi.org/10.5091/plecevo.2019.1612>
- KULIKOVSKIY, M.S.; ANDREEVA, S.A.; GUSEV, E.S.; KUZNETSOVA, I.V. & ANNEKOVA, N.V. (2016a): Molecular phylogeny of monoraphid diatoms and raphe significance in evolution and taxonomy. – *Biological Bulletin* 43: 398–407. DOI: <https://doi.org/10.1134/S1062359016050046>
- KULIKOVSKIY, M.S.; GLUSHCHENKO, A.M.; KUZNETSOVA, I.V. & GENKAL, S.I. (2016b): Identification book of diatoms from Russia. Filigran, – 804 pp., Filigran, Yaroslavl.
- KULIKOVSKIY, M.S.; MALTSEV, Ye.I.; ANDREEVA, S.A.; GLUSHCHENKO, A.M.; GUSEV, E.S.; PODUNAY, YU.A.; LUDWIG, T.V.; TUSSET, E. & KOCIOLEK, J.P. (2019): Description of a new diatom genus *Dorofeyukea* gen. nov. with remarks on phylogeny of the family Stauroneidaceae. – *Journal of Phycology* 55: 173–185. DOI: <https://doi.org/10.1111/jpy.12810>
- LAI, G.G.; PADEDDA, B.M.; WETZEL, C.E.; LUGLIÈ, A.; SECHI, N. & ECTOR, L. (2016): Epilithic diatom assemblages and environmental quality of the Su Gologone karst spring (central–eastern Sardinia, Italy). – *Acta Botanica Croatica* 75: 129–143. DOI: <https://doi.org/10.1515/botcro-2016-0008>
- LAI, G.G.; BURATO, S.; PADEDDA, B.M.; ZORZA, R.; PIZZUL, E.; DELGADO, C.; LUGLIÈ, A. & CANTONATI, M. (2019): Diatom Biodiversity in Karst Springs of Mediterranean Geographic Areas with Contrasting Characteristics: Islands vs Mainland. – *Water* 11: 2602. DOI: <https://doi.org/10.3390/w11122602>
- LEE, K.; CHOI, J.K. & LEE, J.H. (1995): Taxonomic studies on diatoms in Korea. II. Check-list. – *Korean Journal of Phycology* 10: 13–89.
- LEE, J.H. (2012): Algal flora of Korea. Chrysophyta: Bacillariophyceae: Pennales: Raphidinae: Naviculaceae. Freshwater diatoms VI. – pp. 1–56, National Institute of Biological Resources, Incheon.
- LEE, S. S.; TOBIAS, F. A. & VAN DE VIJVER, B. (2013): *Envekadea metzeltinii* sp. nov., a new diatom (Bacillariophyta) species from the subtropical karstic wetlands of the Florida Everglades, USA. – *Phytotaxa* 115: 15–24. DOI: <https://doi.org/10.11646/phytotaxa.115.1.2>
- LINO, C.F.; BOGGIANI, P.C.; CORTESÃO, J.; GODOY, N.M. & KARMANN, I. (1984): Projeto Grutas de Bonito. Diretrizes para um plano de manejo turístico. – 212 pp, SPHAN/MS–TUR, Campo Grande.
- LIU, B.; XIANG, D.; QUAN, S.; LONG, H. & YALUN, M. *Dorofeyukea kotschy* (Grunow) Kulikovskiy, Kociolek, Tusset & T. Ludwig (2020): a new genus and species record from China. – *Acta Botanica Boreali–Occidentalia Sinica* 40: 353–357. DOI: <https://doi.org/10.7606/j.issn.1000-4025.2020.02.0353>
- LOWE, R.; KOCIOLEK, J.P.; YOU, Q.M.; WANG, Q.X. & STEPANEK, J. (2017): Diversity of the diatom genus *Humidophila* in karst areas of Guizhou, China. – *Phytotaxa* 305: 269–284. DOI: <https://doi.org/10.11646/phytotaxa.305.4.3>
- MEDVEDEVA, L.A. & NIKULINA, T.V. (2014): Catalogue of freshwater algae of the southern part of the Russian Far East. Vladivostok: Dalnauka. 271 pp.
- METZELTIN, D. & LANGE–BERTALOT, H. (1998): Tropical Diatoms of South America I: About 700 predominantly rarely known or new taxa representative of the neotropical flora. – In: LANGE–BERTALOT, H. (ed.): *Iconographia Diatomologica* 5. – 695 pp., Koeltz Scientific Books, Königstein, Germany.
- METZELTIN, D. & LANGE–BERTALOT, H. (2002): Diatoms from the “Island Continent” Madagascar. – *Iconographia Diatomologica* 11: 1–286
- MOREIRA–FILHO, H. & VALENTE–MOREIRA, I.M. (1981): Avaliação taxonômica e ecológica das diatomáceas (Bacillariophyceae) epífitas em algas pluricelulares obtidas nos litorais dos Estados do Paraná, Santa Catarina e São Paulo. – *Boletim do Museu Botânico Municipal* 47: 1–17.
- NOVELO, E.; TAVERA, R. & IBARRA, C. (2007): Bacillariophyceae from karstic wetlands in Mexico. – *Bibliotheca Diatomologica* 54: 1–136.
- PATRICK, R.M. (1959): New species and nomenclatural changes in the genus *Navicula* (Bacillariophyceae). – *Proceedings of the Academy of Natural Sciences of Philadelphia*

- 111: 91–108.
- PATRICK, R.M. & REIMER, C.W. (1966): The Diatoms of the United States exclusive of Alaska and Hawaii. Volume 1. Fragilariaceae, Eunotiaceae, Achnanthaceae, Naviculaceae. – Monographs of the Academy of Natural Sciences of Philadelphia 13: 1–688.
- POTAPOVA, M. (2013): The types of 22 *Navicula* (Bacillariophyta) species described by RUTH PATRICK. – Proceedings of the Academy of Natural Sciences of Philadelphia. 162: 1–23. DOI: <http://dx.doi.org/10.1635/053.162.0101>
- ROUND, F.E.; CRAWFORD, R.M. & MANN, D.G. (1990): The diatoms. Biology and morphology of the genera. – 747 pp., Cambridge University Press, Cambridge.
- RUMRICH, U.; LANGE-BERTALOT, H. & RUMRICH, M. (2000): Diatoms of the Andes. From Venezuela to Patagonia/Tierra del Fuego and two additional contributions. – In: LANGE-BERTALOT, H. (ed.): – Iconographia Diatomologica, Annotated Diatom Monographs. Phytogeography–Diversity–Taxonomy, Vol 9. – 673 pp., Koeltz Scientific Books, Königstein, Germany.
- SALLUN FILHO, W.; KARMANN, I. & BOGGIANI, P.C. (2004): Paisagens cársticas da Serra da Bodoquena (MS). – Geologia do Continente Sul-Americano 1: 424–433.
- SALLUN FILHO, W.; KARMANN, I.; BOGGIANI, P.C.; PETRI, S.; DE SOUZA CRISTALLI, P. & UTIDA, G. (2009): A deposição de tufas quaternárias no estado de Mato Grosso do Sul: proposta de definição da formação Serra da Bodoquena. – Geologia USP, Série Científica 9: 47–60. DOI: <https://doi.org/10.5327/Z1519-874x2009000300003>
- SILVA, W.J.; NOGUEIRA, I.S. & SOUZA, M.G.M. (2011): Catálogo de diatomáceas da região Centro-Oeste brasileira. – Iheringia, Série Botânica 66: 61–86.
- SIMONSEN, R. (1974): The diatom plankton of the Indian Ocean expedition of R/V “Meteor” 1964–1965. – Meteor Forsch. Ergebnisse, Reihe D. 19: 1–107.
- STOERMER, E.F.; KREIS, R.G. & ANDRESEN, N.A. (1999): Checklist of Diatoms from the Laurentian Great Lakes. II. – Journal of Great Lakes Research 25: 515–566. DOI: [https://doi.org/10.1016/S0380-1330\(99\)70759-8](https://doi.org/10.1016/S0380-1330(99)70759-8)
- STOYNEVA-GÄRTNER, M.P. & DESCY, J.P. (2020): Checklist of cyanoprokaryotes and algae in the large tropical River Congo (Africa). – Faculty of Biology 104: 18–57.
- TABOADA, M.D.L.Á.; MIRANDA, M.J. & GAVRILOFF, I.J. (2021): Primer relevamiento de la riqueza algal de una laguna de montaña del Noroeste Argentino. – Bonplandia 30: 37–60. DOI: <http://dx.doi.org/10.30972/bon.3014589>.
- TRÁBERT, Z.; BUCZKÓ, K.; BORICS, G.; UDOVIČ, M.G.; KARTHICK, B.; ECTOR, L.; FÖLDI, A.; KISS, K.T. & ÁCS, É. (2019): Biogeography and morphology of a poorly known diatom *Dorofeyukea rostellata* (Hustedt) Kulikovskiy & Kociolek. – Limnetica 38: 743–758. DOI: <https://doi.org/10.23818/limn.38.43>
- TREMARIN, P.I.; FREIRE, E.G.; BERTOLLI, L.M. & LUDWIG, T.A.V. (2009): Catálogo das diatomáceas (Ochrophyta–Diatomeae) continentais do estado do Paraná. – Iheringia, Série Botânica 64: 79–107.
- TUSSET, E.A.; TREMARIN, P.I.; STRAUBE, A. & LUDWIG, T.A. (2017): Morphology of *Adlafia* taxa (Bacillariophyta, Cymbellaceae), with proposition of two new species from Brazil. – Phytotaxa 306: 259–274. DOI: <https://doi.org/10.11646/phytotaxa.306.4.2>
- TUSSET, E.A.; TREMARIN, P.I. & LUDWIG, T.A. (2018): Two new *Stauroneis* species (Bacillariophyta, Stauroneidaceae) from midwestern karstic Brazilian formations. – Phytotaxa 358: 265–277. DOI: <https://doi.org/10.11646/phytotaxa.358.3.4>
- UDOVÍČ, M.G.; KULAŠ, A.; ŠUŠNJARA, M.; ARAPOV, J.; BLANCO, S. & LEVKOV, Z. (2022): *Cymboppleura amicula* stat nov. et. nom. nov. (Bacillariophyceae): a rare diatom species from a karst river in Croatia. – Phytotaxa 532: 139–151. DOI: <https://doi.org/10.11646/phytotaxa.532.2.2>
- WADMARE, N.; BAGHELA, A.; KOCIOLEK, J.P. & KARTHICK, B. (2022): Description and phylogenetic position of three new species of *Stauroneis* Ehrenberg (Bacillariophyceae: Stauroneidaceae) from the Indian Subcontinent. – European Journal of Phycology 57: 48–67. DOI: <https://doi.org/10.1080/09670262.2021.1888390>
- WEIDE, D.M. (2015): *Aulacoseira stevensiae* sp. nov. (Coscinodiscophyceae, Bacillariophyta), a new diatom from Ho Ba Bê, Bac Kan Province, Northern Việt Nam. – Diatom Research 30: 263–268. DOI: <https://dx.doi.org/10.1080/0269249X.2015.1074114>
- WOJTAL, A. (2003): Diatoms of the genus *Gomphonema* Ehr. (Bacillariophyceae) from a karstic stream in the Krakowsko-Czestochowska Upland. – Acta Societatis Botanicorum Poloniae 72: 213–220. DOI: <https://doi.org/10.5586/asbp.2003.028>
- YOU, Q.M.; KOCIOLEK, J.P.; YU, P.; CAI, M.J.; LOWE, R.L. & WANG, Q.X. (2016): A new species of *Simonsenia* from a karst landform, Maolan Nature Reserve, Guizhou Province, China. – Diatom Research 31: 269–275. DOI: <https://doi.org/10.1080/0269249X.2016.1227377>
- YOU, Q.; CAO, Y.; YU, P.; KOCIOLEK, J.P.; ZANG, L.; WU, B.; LOWE, R., & WANG, Q. (2019): Three new subaerial *Achnantheidium* (Bacillariophyta) species from a karst landform in the Guizhou Province, China. – Fottea 19: 138–150. DOI: <https://doi.org/10.5507/fot.2019.005>

Supplementary material

The following supplementary material is available for this article:

Table S1. Morphometric data of the Brazilian population and related *Dorofeyukea* species.

This material is available as part of the online article (<http://fottea.czechphycology.cz/contents>)

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